

Clinical and microbiological evaluation of one-stage full-mouth disinfection: a short-term study

Avaliação clínica e microbiológica do protocolo one-stage full-mouth disinfection: estudo de curto prazo

José Roberto CORTELLI^a, Marcos Vinicius Moreira de CASTRO^a, Rodrigo Dalla Pria BALEJO^a,
Camila Oliveira de ALENCAR^a, Antonio Carlos GARGIONI FILHO^a, Sheila Cavalca CORTELLI^a,
Fernando Oliveira COSTA^b

^aUNITAU – Universidade de Taubaté, Taubaté, SP, Brasil

^bFaculdade de Odontologia, UFMG – Universidade Federal de Minas Gerais, Belo Horizonte, MG, Brasil

Resumo

Introdução: Os pacientes parecem aderir melhor ao tratamento periodontal de curto prazo. Além disso, tratamentos de tempo curto possuem melhor custo-benefício. No entanto, os benefícios associados a este tipo de tratamento ainda requerem investigações adicionais. **Objetivo:** O presente estudo avaliou longitudinalmente indivíduos com periodontite crônica clínica e microbiologicamente tratados pelo protocolo *one-stage full-mouth disinfection*. **Material e método:** Dezesesseis indivíduos ($49,87 \pm 8,22$), que se enquadraram nos critérios de inclusão/exclusão foram incluídos. Um examinador calibrado, avaliou índice de placa e gengival, profundidade de bolsa e nível clínico de inserção pré e pós terapia. Amostras subgengivais coletadas dos cinco sítios periodontais mais doentes estabeleceram a carga bacteriana total e níveis de *P. gingivalis* e *S. oralis* por qPCR. *One-stage full-mouth disinfection* foi realizado com instrumentos manuais associado a gel de clorexidina e solução. Após, os indivíduos utilizaram clorexidina 0,12% para bochecho, duas vezes ao dia, durante os dois meses. Dados foram comparados pelo teste pareado t de Student ($p < 0,05$). **Resultado:** A análise estatística revelou que o tratamento proporcionou melhorias clínicas e microbianas significativas em três meses. A carga bacteriana total evidenciou reduções mais pronunciadas do pré para o pós-tratamento ($p = 0,0001$). Similarmente, *P. gingivalis* e *S. oralis* mostraram redução no pós-tratamento. **Conclusão:** Após 3 meses de monitoramento, indivíduos com periodontite crônica apresentaram melhora clínica e microbiana com o protocolo *one stage full-mouth disinfection*.

Descritores: Periodontite crônica; microbiologia; terapia; *Porphyromonas gingivalis*; *Streptococcus oralis*.

Abstract

Introduction: Patients seem to adhere better to short-term periodontal treatment schemes. Besides, time-reduced treatments are more cost-effective. However, the degree of benefits related to this type of treatment still requires additional investigations. **Aim:** The present short-term study evaluated clinical and microbiological outcomes, from baseline to 3-months, of chronic periodontitis subjects treated by the one-stage full-mouth disinfection protocol. **Material and method:** Sixteen chronic periodontitis subjects (mean-age 49.87 ± 8.22) who met inclusion/exclusion criteria were included. A calibrated examiner measured whole-mouth plaque and gingival indices, periodontal pocket depth and clinical attachment level at baseline and at 3-months. Subgingival samples were also collected from the 5 most diseased periodontal sites to determine total bacterial load and levels of *P. gingivalis* and *S. oralis* by real time qPCR. Periodontal treatment consisted of full-mouth manual debridement plus wide intraoral use of chlorhexidine in gel and solution. Additionally, after debridement, individuals rinsed 0.12% chlorhexidine at home twice a day for the following 2 months. Data monitored were compared by paired Student-t test ($p < 0.05$). **Result:** Statistical analysis revealed that, in general, one-stage full-mouth disinfection treatment provided significant clinical and microbiological improvements at 3-months. Total bacterial load showed one of the most pronounced reductions from baseline to 3-months ($p = 0.0001$). Also, subgingival levels *P. gingivalis* and *S. oralis* reduced overtime. **Conclusion:** After a short period of monitoring, chronic periodontitis subjects showed clinical and microbial improvements following one-stage full-mouth disinfection treatment.

Descriptors: Chronic periodontitis; microbiology; therapy; *Porphyromonas gingivalis*; *Streptococcus oralis*.

INTRODUCTION

Periodontal diseases are one of the most common in the oral cavity and have an infectious nature triggered by pathogenic microorganisms that first lead to gingivitis and may develop to periodontitis and, if not treated, to tooth loss^{1,2}. Gingivitis is clinically verified by spontaneous or provoked (depending on its severity) bleeding. Periodontitis is characterized by loss of attachment in the conjunctive tissues followed by alveolar bone destruction and consequent formation of periodontal pockets^{1,2}.

Among several bacterial species capable of colonizing the human oral cavity, *Porphyromonas gingivalis*, a Gram-negative anaerobe colonizes distinct niches and is described as one of the major pathogens associated with periodontal breakdown³. In relation to the total bacterial counts, levels of *P. gingivalis* are percentually higher in periodontally diseased sites than in healthy sites⁴ while *Streptococcus oralis* is known as the predominant colonizer in the early stage of dental plaque biofilm formation⁵.

Because of the microbial nature of periodontal disease its prevention requires reducing biofilm⁶ and educating individuals to a good level of oral hygiene⁷. In despite of periodontal disease can be treated successfully by means of both mechanical non-surgical and surgical therapy, periodontal maintenance therapy is a crucial factor for the success of periodontal treatment⁸.

After a long time of quadrant treatment, the concept of one-stage full mouth disinfection⁹ emerged to avoid transmission of pathogenic microorganisms from not treated periodontal pockets to those recently debrided and thus in the healing process. The original protocol proposed by Quirynen's group adopted combined mechanical and chemical procedures to eliminate plaque and/or calculus from teeth and other oral microbial reservoirs such as tongue and tonsils, thus promoting full-mouth disinfection within a 24-hour period. Periodontal pockets were irrigated by chlorhexidine rinse which was also used daily at home as an additional preventive measure to control biofilm formation. Some of the benefits of this protocol were related to a potential stimulus of the immunological response and a better cost-benefit relation⁹.

Therefore, this present short-term study evaluated clinical and microbiological effectiveness of the one-stage full-mouth disinfection protocol in a sample of chronic periodontitis subjects using a three-month longitudinal design.

MATERIAL AND METHOD

The subjects included in the current study signed an informed consent form, which had been previously approved by the Institutional Committee on Research Involving Human Subjects of the University of Taubate (Protocol 521/10).

The study population was composed of sixteen generalized chronic periodontitis individuals registered in the screening periodontal program of the University of Taubate, school of dentistry in the year of 2012. All participants both genders

were 18 or older, had at least 20 natural teeth and had chronic periodontitis according to the criteria established¹⁰.

1. Clinical Periodontal Examinations

Participants were submitted to a complete periodontal examination, in three times, [1] first to determine a periodontal diagnosis, [2] second to establish baseline data and [3] third, 90 days after the one stage full-mouth disinfection protocol intervention to assess treatment effectiveness. Periapical and or panoramic radiographs were taken only in the first periodontal examination [1].

Probing depth (PD), clinical attachment level (CAL), plaque index¹¹ (PI) and gingival index¹² (GI) were taken from all teeth, except for third molars, with the help of a manual periodontal probe (PCPUNC 15 Hu-friedy Mfg Co Inc. Chigago IL). One trained and calibrated examiner, blinded to type of treatment, conducted all clinical measurements. Baseline data analysis was performed to determine if the intraexaminer reliability was calibrated. Using kappa statistics (*K*) for the categorical clinical measurement variables, such as periodontal probing depth and clinical attachment level, the standard error of these measurements was monitored. The examiner's clinical measurement technique was considered calibrated if the standard error for the measurements was ≤ 0.8 and a *K* value ranged between 0.8 and 0.95. The reproducibility of the intraexaminer measurements was recalculated prior to the final clinical exams.

Debridement procedures were carried in a single stage (within 24 hours) with Gracey and McCall cures and Hirschfield files. The process was divided in two appointments (60 minutes each) in two consecutive days. In the beginning of each session subjects rinsed 20 ml 0.12% chlorhexidine (CHX) solution for 30 seconds (the last 10 seconds were of gargling), and in the end of each session there was tongue brushing for 1 minute with a 1% CHX gel and, again, rinsing for 30 seconds. Additionally, individuals were instructed to rinse 20 ml 0.12% CHX for 30 seconds twice a day for 60 consecutive days after the single stage debridement.

2. Sampling for Microbial Analysis and qPCR Reaction

Subgingival samples were collected based on the methodology previously used by our group¹³. More specifically, subgingival samples were collected from mesio-buccal aspect of the five teeth showing more evidence of periodontal disease (preferably showing higher values of probing depth, clinical attachment loss and gingival inflammation) using sterile paper points, number 30, inserted into the depth of the pocket after the removal of supragingival plaque using sterile cures. Sixty seconds after placement in the pocket, paper points were immediately inserted in a microtube and kept on ice. The bacterial cells in the microtubes were dispersed using a vortex mixer at maximum speed for one minute, and the resulting bacterial suspension was saved in a freezer at -80°C until laboratorial processing.

Genomic DNA (gDNA) was extracted and purified from the pellet using PureLink® Genomic DNA Mini Kit (Life Technologies, Carlsbad, CA, USA) according to manufacturer's specifications.

The quantification of total number of bacterial cells, *P. gingivalis* and *S. oralis* was carried out by qPCR using TaqMan assay (TaqMan® Universal PCR Master Mix II, Life Technologies) with specific set of primers/probes (Table 1) in an ABI 7500 Fast Real Time PCR System® (Life Technologies) following manufacturer's instructions in 20ul reactions. The qPCR conditions were: 50 °C for 2 minutes, 95 °C for 10 minutes, 40 cycles of 95 °C for 15 seconds and 60 °C for 1 minute.

The absolute quantification of the target organisms were determined by the plotting of the cycle threshold (Ct) value obtained from each clinical sample against a standard curve generated with known concentration of gDNA of reference bacterial strains (Table 1) in 10-fold serial dilutions (10^2 - 10^7 cells). Negative control (purified PCR-grade water instead of the DNA template) was included in all PCR reactions.

3. Statistical Analysis

A statistical analysis was performed comparing data from baseline to 3-months. Clinical and microbiological data was statistically analyzed using paired-Student t test. The level

of statistical significance adopted was 95% ($\alpha=0.05$) and the software was SPSS 13.0 (IBM® SPSS® Statistics).

RESULT

The present study included 16 individuals: 8 women and 8 men (49.87 ± 8.22 mean age). The evaluation of clinical PD, CAL, PI, and GI showed that all parameters significantly improved after treatment (Table 2). In addition, it was observed that total bacterial load (universal), *P. gingivalis* and *S. oralis* showed a statistically significant ($p<0.05$) reduction after periodontal treatment (Figure 1).

DISCUSSION

Traditionally, periodontal debridement procedures are made in quadrants or sextants with regular intervals of one or two weeks. It is known that the clinical success of this type of treatment happens mainly because of the reduction of periodontal pathogens generally accompanied by the increase of beneficial bacteria¹⁶ and the subsequent establishment of a healthy microbiota. However, these microbial changes could be impaired in individuals with more severe periodontal disease, since they are more susceptible to intra-oral cross contamination involving the variety of aerobic

Table 1. Primers/Probes and reference stains used for microbial quantification by qPCR

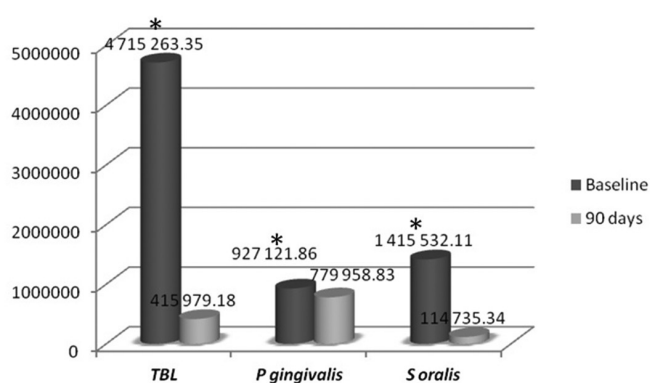
Primers/Probe	Reference	Reference stain
Universal		
Forward: 5'-TGGAGCATGTGGTTAATTCGA-3'	Nonnenmacher et al. ¹⁴ (2004)	
Reverse: 5'-TGCGGGACTTAACCCAACA-3'	Nonnenmacher et al. ¹⁴ (2004)	<i>Escherichia coli</i> (ATCC 25922)
Probe: 5'-CACGAGCTGACGACA(AG)CCATGCA-3'	Nonnenmacher et al. ¹⁴ (2004)	
<i>P. gingivalis</i>		
Forward: 5'-ACCTTACCCGGGATTGAAATG-3'	Saito et al. ¹⁵ (2011)	
Reverse: 5'-CAACCATGCAGCACCTACATAGAA-3'	Saito et al. ¹⁵ (2011)	<i>P. gingivalis</i> (W83)
Probe: 5'-ATGACTGATGGTGA AACCGTCTTCCCTTC-3'	Saito et al. ¹⁵ (2011)	
<i>S. oralis</i>		
Forward: 5'-TTGGCTCAATCCCTTTGAC-3'	Designed for this study*	
Reverse: 5'-GTCCAACAAGCCACCACTT-3'	Designed for this study*	<i>S. oralis</i> (ATCC 10557)
Probe: 5'-ACAACATATCAACAGGCGCA-3'	Designed for this study*	

*Designed using Primer3 software online version (v. 0.4.0). NCBI Blast database was used to check Primer/Probe specificity.

Table 2. Distribution of clinical periodontal parameters before (baseline) and 90 days after periodontal treatment

	PD	CAL	PI	GI
Baseline	3.72 ± 0.95	5.01 ± 1.92	0.43 ± 0.23	0.35 ± 0.34
90 days	2.13 ± 0.76	3.54 ± 1.82	0.24 ± 0.27	0.13 ± 0.17
<i>p</i> value	0.0321	0.0138	0.0295	0.0341

Paired Student-t test, Wilcoxon ($p<0.05$).



* - Statistically significant difference ($p < 0.05$) paired Student-t test.

Figure 1. Distribution of the final number of copies before and after treatment for universal primer, *P. gingivalis* and *S. oralis*

and anaerobic bacteria¹⁷ which can translocate from not treated sites to those recently treated. In this context, the concept of one-stage full mouth disinfection emerges and aims mainly at minimizing transmission of pathogenic microorganisms from the periodontal pockets still not treated to those already treated⁹.

Chlorhexidine is an antimicrobial agent widely used in Dentistry. Although in this protocol CHX had demonstrated good results, authors felt like it was necessary to check the degree of benefit brought by the product^{18,19}. Therefore, we decided that would be interesting to contribute to clarify doubts left by other studies such as that of which concluded that the greatest benefit derived from intense mechanical therapy carried within 24 hours²⁰. In a posterior study, however, the same group reported controversial results after observing that the prolonged use of CHX brought beneficial effects to the protocol. During explanation of potential risks and benefits of this study, subjects were informed about possible side effects related to CHX. Patients did not complain about side effects although examiner was able to identify teeth staining.

Individuals included in this study showed a microbial profile with high copies of bacterial DNA (see Figure 1) verified by q-PCR in baseline, so the adoption of the 24 hour treatment seemed convenient and beneficial, since it could minimize the chance of intra-oral transmissibility. Such hypothesis was verified when a drastic bacterial reduction was observed by the final reduced total bacterial load after therapy. This finding corroborates the study which affirmed that one-stage full mouth protocol immediately decrease the number of bacteria in mild to advanced periodontitis. This reduction will positively affect the

recolonization process due to the slowing of supragingival and consequently subgingival biofilm formation²¹.

P. gingivalis, is one of the many bacteria colonizing the mouth, found in different sites such as periodontal pockets, oral mucosa and tongue dorsum^{22,23}. It is a Gram-negative microorganism and an important etiologic agent of periodontal disease. Colonization by this pathogen results in tissue lesion due to the exacerbated production of a variety of virulence factors. Besides, this bacterium seems to have a significant role in the progression of chronic periodontitis^{24,25} and, in high levels, is usually related to deep periodontal pockets²⁶. Our study showed that, after therapy, there was a statistically significant reduction in the levels of *P. gingivalis* in comparison to baseline. Total bacterial reduction associated with the decrease in the levels of *P. gingivalis* may easily be connected to the improvement of clinical variables observed in this study. All parameters – PD, CAL, PI, PG – showed statistically significant improvement after therapy. It is known that some clinical periodontal parameters respond more quickly following treatment while others only manifest improvement or worsening later.

S. oralis is known as the predominant colonizer in the early stage of dental plaque biofilm formation²⁷. Primary colonizers alter the surface not only by their physical presence but also they are likely to represent a new “surface-attached” phenotype with distinct metabolic activity and surface properties, thus altering their surroundings and creating new niches for other bacteria to colonize²⁸. This specie seems to be less pathogenic for periodontitis, and our results showed that the prevalence was markedly low in periodontitis patients, like *P. gingivalis*. We assumed that the considerable amount of *S. oralis* could be eliminated from the periodontal pockets by initial treatments, and its prevalence in subgingival plaque would decrease in the present study.

After a short-term period of monitoring results obtained in this study have lead us to conclude that, one-stage full mouth disinfection protocol can be a treatment option to chronic periodontitis subjects. In addition, our study demonstrated that the adjunctive daily use of an antimicrobial agent to mechanical plaque control could result in a retarded subgingival recolonization by periodontal pathogens in examined population.

ACKNOWLEDGEMENTS

This study was sponsored by São Paulo Research Foundation - FAPESP – São Paulo/SP – Brazil (Process # 2010/19079-8).

REFERENCES

1. Løe H, Anerud A, Boysen H, Morrison E. Natural history of periodontal disease in man. Rapid, moderate and no loss of attachment in Sri Lankan laborers 14 to 16 years of age. J Clin Periodontol. 1986; 13: 431-40. PMID:3487557. <http://dx.doi.org/10.1111/j.1600-051X.1986.tb01487.x>
2. Lindhe J, Karring T, Lang NP. Tratado de periodontia clínica e implantodontia oral. Rio de Janeiro: Guanabara Koogan; 2005.
3. Zambon JJ. Periodontal diseases: microbial factors. Ann Periodontol. 1996; 1: 879-925. PMID:9118283. <http://dx.doi.org/10.1902/annals.1996.1.1.879>

4. Kumar PS, Leys EJ, Bryk JM, Martinez FJ, Moeschberger ML, Griffen AL. Changes in periodontal health status are associated with bacterial community shifts as assessed by quantitative 16S cloning and sequencing. *J Clin Microbiol.* 2006; 44: 3665-73. PMID:17021095 PMCID:PMC1594761. <http://dx.doi.org/10.1128/JCM.00317-06>
5. Li J, Helmerhorst EJ, Leone CW, Troxler RF, Yaskell T, Haffajee AD, et al. Identification of early microbial colonizers in human dental biofilm. *J Appl Microbiol.* 2004; 97: 1311-8. PMID:15546422. <http://dx.doi.org/10.1111/j.1365-2672.2004.02420.x>
6. Adriaens PA, Adriaens LM. Effects of nonsurgical periodontal therapy on hard and soft tissues. *Periodontol 2000.* 2004 Oct; 36(1): 121-45. PMID:15330946. <http://dx.doi.org/10.1111/j.1600-0757.2004.03676.x>
7. Suomi, JD, Greeme JC, Vermillion JR, Doyle J, Chang JJ, Leatherwood EC. The effect of controlled oral hygiene procedures on the progression of periodontal disease in adults: results after third and final year. *J Periodontol.* 1971; 42(3): 152-60. PMID:4396693. <http://dx.doi.org/10.1902/jop.1971.42.3.152>
8. Costa FO, Santuchi CC, Lages EJ, Cota LO, Cortelli SC, Cortelli JR, et al. Prospective study in periodontal maintenance therapy: comparative analysis between academic and private practices. *J Periodontol.* 2012; 83(3):301-11. PMID:21780903. <http://dx.doi.org/10.1902/jop.2011.110101>
9. Quirynen M, Bollen CM, Vandekerckhove BN, Dekeyser C, Papaioannou W, Eysen H. Full-vs. partial-mouth disinfection in the treatment of periodontal infections: short-term clinical and microbiological observations. *J Dent Res.* 1995; 74: 1459-67. PMID:7560400. <http://dx.doi.org/10.1177/00220345950740080501>
10. Armitage GC. Development of a classification system for periodontal diseases and conditions. *Ann Periodontol.* 1999; 4: 1-6. PMID:10863370. <http://dx.doi.org/10.1902/annals.1999.4.1.1>
11. Silness J, Løe H. Periodontal disease in pregnancy II. Correlation between oral hygiene and periodontal condition. *Acta Odontol Scand.* 1964; 22: 121-35. PMID:14158464. <http://dx.doi.org/10.3109/00016356408993968>
12. Løe H, Silness J. Periodontal disease in pregnancy. I. Prevalence and severity. *Acta Odontol Scand.* 1963; 21: 533-51. PMID:14121956. <http://dx.doi.org/10.3109/00016356309011240>
13. Cortelli JR, Cortelli SC, Jordan S, Haraszthy VI, Zambon JJ. Prevalence of periodontal pathogens in Brazilians with aggressive or chronic periodontitis. *J Clin Periodontol.* 2005; 32: 860-6. PMID:15998269. <http://dx.doi.org/10.1111/j.1600-051X.2005.00777.x>
14. Nonnenmacher C, Dalpke A, Mutters R, Heeg K. Quantitative detection of periodontopathogens by real-time PCR. *J Microbiol Methods.* 2004 Oct; 59(1): 117-25. PMID:15325758. <http://dx.doi.org/10.1016/j.mimet.2004.06.006>
15. Saito T, Inagaki S, Sakurai K, Okuda K, Ishihara K. Exposure of *P. gingivalis* to noradrenaline reduces bacterial growth and elevates ArgX protease activity. *Arch Oral Biol.* 2011 Mar; 56(3): 244-50. PMID:20970116. <http://dx.doi.org/10.1016/j.archoralbio.2010.09.014>
16. Ali RW, Lie T, Skaug N. Early effects of periodontal therapy on the detection frequency of four putative periodontal pathogens in adults. *J Periodontol.* 1992; 63: 540-7. PMID:1625154. <http://dx.doi.org/10.1902/jop.1992.63.6.540>
17. Tan BT, Mordan NJ, Embleton J, Pratten J, Galgut PN. Study of bacterial viability within human supragingival dental calculus. *J Periodontol.* 2004; 75: 23-9. PMID:15025213. <http://dx.doi.org/10.1902/jop.2004.75.1.23>
18. Bollen CM, Vandekerckhove BN, Papaioannou W, Van Eldere J, Quirynen M. Full- versus partial-mouth disinfection in the treatment of periodontal infections. A pilot study: long term microbiological observations. *J Clin Periodontol.* 1996; 23: 960-70. PMID:8915027. <http://dx.doi.org/10.1111/j.1600-051X.1996.tb00519.x>
19. Mongardini C, Van Steenberghe D, Dekeyser C, Quirynen M. One stage full- versus partial-mouth disinfection in the treatment of chronic adult or generalized early-onset periodontitis. I. Long-term clinical observations. *J Periodontol.* 1999; 70: 632-45. PMID:10397519. <http://dx.doi.org/10.1902/jop.1999.70.6.632>
20. Quirynen M, Mongardini C, De Soete M, Pauwels M, Coucke W, Van Eldere J, et al. The role of chlorhexidine in the one-stage full-mouth disinfection treatment of patients with advanced adult periodontitis. Long-term clinical and microbiological observations. *J Clin Periodontol.* 2000; 27: 578-89. PMID:10959784. <http://dx.doi.org/10.1034/j.1600-051x.2000.027008578.x>
21. Sekino S, Ramberg P, Uzel NG, Socransky SS, Lindhe J. The effect of a chlorhexidine regimen on de novo plaque formation. *J Clin Periodontol.* 2004; 31: 609-14. PMID:15257736. <http://dx.doi.org/10.1111/j.1600-051X.2004.00526.x>
22. Van Steenberghe TJ, Petit MD, Scholte LH, van der Welden U, de Graff J. Transmission of *Porphyromonas gingivalis* between spouses. *J Clin Periodontol.* 1993; 20: 340-5. PMID:8388896. <http://dx.doi.org/10.1111/j.1600-051X.1993.tb00370.x>
23. Cortelli JR, Aquino DR, Cortelli SC, Nobre Franco GC, Fernandes CB, Roman-Torres CV, et al. Detection of periodontal pathogens in oral mucous membranes of edentulous individuals. *J Periodontol.* 2008 Oct; 79(10): 1962-5. PMID:18834252. <http://dx.doi.org/10.1902/jop.2008.080092>
24. Lamont RJ, Jenkinson HF. Life below the gum line: pathogenic mechanisms of *Porphyromonas gingivalis*. *Microbiol Mol Biol.* 1998; 62: 1244-63.
25. Dzink JL, Socransky SS, Haffajee AD. The predominant cultivable microbiota of active and inactive lesions of destructive periodontal diseases. *J Clin Periodontol.* 1988; 15: 316-23. PMID:3292595. <http://dx.doi.org/10.1111/j.1600-051X.1988.tb01590.x>
26. Van Winkelhoff AJ, Van der Velden U, Winkel EG, de Graff J. Black-pigmented *Bacteroides* and motile organisms in individuals with and without periodontal breakdown. *J Periodontol Res.* 1986; 21: 434-8. PMID:2942671. <http://dx.doi.org/10.1111/j.1600-0765.1986.tb01477.x>

27. Haffajee AD, Uzel NG, Arguello EI, Torresyap G, Guerrero DM, Socransky SS. Clinical and microbiological changes associated with the use of combined antimicrobial therapies to treat “refractory” periodontitis. *J Clin Periodontol.* 2004; 31: 869-77. PMID:15367191. <http://dx.doi.org/10.1111/j.1600-051X.2004.00573.x>
28. Davey ME, Costerton JW. Molecular genetics analyses of biofilm formation in oral isolates. *Periodontol* 2000. 2006; 42: 13-26. PMID:16930303. <http://dx.doi.org/10.1111/j.1600-0757.2006.00052.x>

CONFLICTS OF INTERESTS

The authors declare no conflicts of interest.

CORRESPONDING AUTHOR

José Roberto Cortelli
Departamento de Odontologia, UNITAU – Universidade de Taubaté, Rua Expedicionário Ernesto Pereira, 110, 12020-330
Taubaté - SP, Brasil
e-mail: jrcortelli@uol.com.br

Received: June 06, 2013
Accepted: August 12, 2013